

# Package ‘martini’

May 15, 2025

**Type** Package

**Title** GWAS Incorporating Networks

**Version** 1.29.0

**Description** martini deals with the low power inherent to GWAS studies by using prior knowledge represented as a network. SNPs are the vertices of the network, and the edges represent biological relationships between them (genomic adjacency, belonging to the same gene, physical interaction between protein products). The network is scanned using SConES, which looks for groups of SNPs maximally associated with the phenotype, that form a close subnetwork.

**License** GPL-3

**LazyData** TRUE

**Imports** igraph (>= 1.0.1), Matrix, memoise (>= 2.0.0), methods (>= 3.3.2), Rcpp (>= 0.12.8), snpStats (>= 1.20.0), stats, utils,

**Suggests** biomaRt (>= 2.34.1), circlize (>= 0.4.11), STRINGdb (>= 2.2.0), httr (>= 1.2.1), IRanges (>= 2.8.2), S4Vectors (>= 0.12.2), knitr, testthat, readr, rmarkdown

**Depends** R (>= 4.0)

**LinkingTo** Rcpp, RcppEigen (>= 0.3.3.5.0)

**RoxygenNote** 7.2.3

**Encoding** UTF-8

**biocViews** Software, GenomeWideAssociation, SNP, GeneticVariability, Genetics, FeatureExtraction, GraphAndNetwork, Network

**VignetteBuilder** knitr

**URL** <https://github.com/hclimente/martini>

**BugReports** <https://github.com/hclimente/martini/issues>

**git\_url** <https://git.bioconductor.org/packages/martini>

**git\_branch** devel

**git\_last\_commit** 1253ale

**git\_last\_commit\_date** 2025-04-15

**Repository** Bioconductor 3.22

**Date/Publication** 2025-05-15

**Author** Hector Climente-Gonzalez [aut, cre] (ORCID:  
 <<https://orcid.org/0000-0002-3030-7471>>),  
 Chloe-Agathe Azencott [aut] (ORCID:  
 <<https://orcid.org/0000-0003-1003-301X>>)

**Maintainer** Hector Climente-Gonzalez <[hector.climente@riken.jp](mailto:hector.climente@riken.jp)>

## Contents

arrange_covars	3
calculateE	3
calculateG	4
check_installed	4
connect_biomart	5
get_adjacency	5
get_GI_network	6
get_GM_network	7
get_grid	8
get_GS_network	8
get_gxg	9
get_gxg_biogrid	9
get_gxg_string	10
get_snp_modules	10
group_snps	11
gwas2bed	11
is_coherent	12
ldweight_edges	12
maxflow	13
mget_gxg_biogrid	13
mget_gxg_string	14
mincut	14
mincut.cv	15
mincut_c	15
minigwas	16
minippi	16
minisnpMapping	17
organism_id2name	17
permute_snpMatrix	17
plot_ideogram	18
sanitize_map	18
sanitize_snpMapping	19
scones	19
scones.cv	20
scones.cv_	21
scones_	22
score_fold	23
search_cones	23
sigmod	25
sigmod.cv	26
sigmod.cv_	27
sigmod_	28
simulate_causal_snps	28

<code>arrange_covars</code>	3
<code>simulate_phenotype</code>	29
<code>snp2ensembl</code>	30
<code>snp_test</code>	31
<code>subnet</code>	31
<code>subset_snpMatrix</code>	32
<code>subvert</code>	32
<code>wrap_Xy</code>	33
<b>Index</b>	<b>34</b>

---

<code>arrange_covars</code>	<i>Prepare covariates for scones</i>
-----------------------------	--------------------------------------

---

**Description**

Prepares de covariates data.frame for the functions used in scones, like `single_snp_association` or `score_folds`.

**Usage**

```
arrange_covars(gwas, covars)
```

**Arguments**

<code>gwas</code>	A SnpMatrix object with the GWAS information.
<code>covars</code>	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.

**Value**

The covars data.frame, with the rows in the same order as gwas.

---

<code>calculateE</code>	<i>Calculate the environmental component of the phenotype</i>
-------------------------	---------------------------------------------------------------

---

**Description**

Calculates the environmental component of the phenotype using the variance in the genetic component.

**Usage**

```
calculateE(G, h2)
```

**Arguments**

<code>G</code>	The genetic component of the phenotype.
<code>h2</code>	The heritability.

**Value**

A vector with the environmental component of each sample.

---

calculateG	<i>Calculate the genetic component of the phenotype</i>
------------	---------------------------------------------------------

---

**Description**

Calculates the genetic component of the phenotype from a genotype.

**Usage**

```
calculateG(effectSize, X, model)
```

**Arguments**

effectSize	A vector with the effect size of each SNP.
X	Genotypes in a numeric matrix, where each row is a sample and each column a SNP.
model	Genetic model to assume.

**Value**

A vector with the genetic component of each sample.

---

check_installed	<i>Check package is installed</i>
-----------------	-----------------------------------

---

**Description**

Checks if a package is installed, launches an error if it is not.

**Usage**

```
check_installed(pkgs, fn = "This function")
```

**Arguments**

pkgs	Character vector with the names of the packages.
fn	Function calling the check.

**Value**

The package is loaded into the namespace.

**Examples**

```
martini:::check_installed(c("martini"))  
## Not run: martini:::check_installed("martinid")
```

---

connect_biomart	<i>Open a biomaRt connection</i>
-----------------	----------------------------------

---

**Description**

Opens a biomaRt connection for the relevant species.

**Usage**

```
connect_biomart(organism)
```

**Arguments**

organism	String containing the ensembl species name (e.g. hsapiens for human)
----------	----------------------------------------------------------------------

---

get_adjacency	<i>Compute Laplacian matrix</i>
---------------	---------------------------------

---

**Description**

Compute Laplacian matrix

**Usage**

```
get_adjacency(gwas, net)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.

**Value**

A Laplacian matrix.

---

get\_GI\_network                      *Get gene-interaction network.*

---

### Description

Creates a network of SNPs where each SNP is connected as in the **GM** network and, in addition, to all the other SNPs pertaining to any interactor of the gene it is mapped to. Corresponds to the gene-interaction (GI) network described by Azencott et al.

### Usage

```
get_GI_network(
  gwas,
  organism = 9606,
  snpMapping = snp2ensembl(gwas, organism),
  ppi = get_gxg("biogrid", organism, flush),
  col_ppi = c("gene1", "gene2"),
  col_genes = c("snp", "gene"),
  flush = FALSE
)
```

### Arguments

gwas	A SnpMatrix object with the GWAS information.
organism	Tax ID of the studied organism. The default is 9606 (human).
snpMapping	A data.frame informing how SNPs map to genes. It contains minimum two columns: SNP id and a gene it maps to. Each row corresponds to one gene-SNP mapping. Unless column names are specified using col_genes, involved columns must be named 'snp' and 'gene'.
ppi	A data.frame describing protein-protein interactions with at least two columns. Gene ids must be the contained in snpMapping. Unless column names are specified using col_ppi, involved columns must be named gene1 and gene2.
col_ppi	Optional, length-2 character vector with the names of the two columns involving the protein-protein interactions.
col_genes	Optional, length-2 character vector with the names of the two columns involving the SNP-gene mapping. The first element is the column of the SNP, and the second is the column of the gene.
flush	Remove cached results? Boolean value.

### Value

An igraph network of the GI network of the SNPs.

### References

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

## Examples

```
get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
```

---

get_GM_network	<i>Get gene membership network.</i>
----------------	-------------------------------------

---

## Description

Creates a network of SNPs where each SNP is connected as in the [GS](#) network and, in addition, to all the other SNPs pertaining to the same gene. Corresponds to the gene membership (GM) network described by Azencott et al.

## Usage

```
get_GM_network(  
  gwas,  
  organism = 9606,  
  snpMapping = snp2ensembl(gwas, organism),  
  col_genes = c("snp", "gene")  
)
```

## Arguments

gwas	A SnpMatrix object with the GWAS information.
organism	Tax ID of the studied organism. The default is 9606 (human).
snpMapping	A data.frame informing how SNPs map to genes. It contains minimum two columns: SNP id and a gene it maps to. Each row corresponds to one gene-SNP mapping. Unless column names are specified using col_genes, involved columns must be named 'snp' and 'gene'.
col_genes	Optional, length-2 character vector with the names of the two columns involving the SNP-gene mapping. The first element is the column of the SNP, and the second is the column of the gene.

## Value

An igraph network of the GM network of the SNPs.

## References

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

## Examples

```
get_GM_network(minigwas, snpMapping = minisnpMapping)
```

---

get\_grid *Parse scones.cv settings*

---

### Description

Creates a list composed by all scones.cv settings, with the values provided by the user, or the default ones if none is provided.

### Usage

```
get_grid(c = numeric(), etas = numeric(), lambdas = numeric())
```

### Arguments

c	Numeric vector with the association scores of the SNPs. Specify it to automatically an appropriate range of etas and lambdas.
etas	Numeric vector with the etas to explore in the grid search. If omitted, it's automatically created based on the association scores.
lambdas	Numeric vector with the lambdas to explore in the grid search. If omitted, it's automatically created based on the association scores.

### Value

A list of scones.cv settings.

### Examples

```
martini:::get_grid(etas = c(1,2,3), lambdas = c(4,5,6))
martini:::get_grid(c = c(1,10,100))
```

---

get\_GS\_network *Get genomic sequence network*

---

### Description

Creates a network of SNPs where each SNP is connected to its adjacent SNPs in the genome sequence. Corresponds to the genomic sequence (GS) network described by Azencott et al.

### Usage

```
get_GS_network(gwas)
```

### Arguments

gwas	A SnpMatrix object with the GWAS information.
------	-----------------------------------------------

### Value

An igraph network of the GS network of the SNPs.



## References

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

## Examples

```
get_GS_network(minigwas)
```

---

get_gxg	<i>Get gene interactions</i>
---------	------------------------------

---

## Description

Wrapper for the different functions to get gene-gene interactions. Supports cached results.

## Usage

```
get_gxg(db, organism, flush)
```

## Arguments

db	String containing the database to obtain the gene-gene interactions from. Possible values: 'biogrid', 'string'.
organism	Tax ID of the studied organism. The default is 9606 (human).
flush	Remove cached results? Boolean value.

## Value

A data.frame with two columns with pairs of interacting proteins.

---

get_gxg_biogrid	<i>Get BioGRID protein-protein interactions.</i>
-----------------	--------------------------------------------------

---

## Description

Get all protein-protein interactions for an organism from BioGRID.

## Usage

```
get_gxg_biogrid(organism = 9606)
```

## Arguments

organism	Tax ID of the studied organism. The default is 9606 (human).
----------	--------------------------------------------------------------

## Value

A data.frame with two columns with pairs of interacting proteins.

**Examples**

```
# download dog interactions
## Not run: martini::get_gxg_biogrid(9615)
```

---

```
get_gxg_string          Get STRING protein-protein interactions.
```

---

**Description**

Get all protein-protein interactions for an organism from STRING. It uses a score cut-off of 400.

**Usage**

```
get_gxg_string(organism = 9606)
```

**Arguments**

organism            Tax ID of the studied organism. The default is 9606 (human).

**Value**

A data.frame with two columns with pairs of interacting proteins.

**Examples**

```
# download frog interactions
## Not run: martini::get_gxg_string(8364)
```

---

```
get_snp_modules        Return groups of interconnected SNPs.
```

---

**Description**

Find modules composed by interconnected SNPs.

**Usage**

```
get_snp_modules(gwas, net)
```

**Arguments**

gwas                A SnpMatrix object with the GWAS information.  
net                 An igraph network that connects the SNPs.

**Value**

A list with the modules of selected SNPs.

**Examples**

```
## Not run:
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
cones <- search_cones(minigwas, gi)
martini::get_snp_modules(cones, gi)

## End(Not run)
```

---

group_snps	<i>Groups nearby SNPs</i>
------------	---------------------------

---

**Description**

Groups SNPs closer than a specific threshold of distance.

**Usage**

```
group_snps.bed, chr_col, pos_col, threshold)
```

**Arguments**

bed	data.frame containing at least two properties (chromosome and position) of a set of SNPs.
chr_col	Name of the column containing the SNP chromosome.
pos_col	Name of the column containing the SNP position.
threshold	Maximum distance to group two SNPs group.

**Value**

A data.frame in bed format, with the same dimensions as the original, but with the groups.

---

gwas2bed	<i>Converts a MAP data.frame to a BED data.frame</i>
----------	------------------------------------------------------

---

**Description**

Takes a map file and:

- column 1: Used as the chromosome column in the BED file.
- column 4: Used as start and end in the BED data.frame (as we work with SNPs).

**Usage**

```
gwas2bed(gwas)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
------	-----------------------------------------------

**Value**

A BED data.frame.

---

is_coherent	<i>Check inner coherence of GWAS dataset</i>
-------------	----------------------------------------------

---

**Description**

Checks that the different data structures have the SNPs in the same order.

**Usage**

```
is_coherent(gwas)
```

**Arguments**

gwas                    A SnpMatrix object with the GWAS information.

**Value**

TRUE if the GWAS dataset is coherent. Else, raises an error.

**Examples**

```
martini:::is_coherent(minigwas)
```

---

ldweight_edges	<i>Include linkage disequilibrium information in the network.</i>
----------------	-------------------------------------------------------------------

---

**Description**

Include linkage disequilibrium information in the SNP network. The weight of the edges will be lower the higher the linkage is.

**Usage**

```
ldweight_edges(net, ld, method = "inverse")
```

**Arguments**

net                    A SNP network.

ld                    A dsCMatrix or dgCMatrix containing linkage disequilibrium measures, like the output of `ld`.

method                How to incorporate linkage-disequilibrium values into the network.

**Value**

An copy of net where the edges weighted according to linkage disequilibrium.

**Examples**

```
ld <- snpStats::ld(minigwas[['genotypes']], depth = 2, stats = "R.squared")
# don't weight edges for which LD cannot be calculated
ld[is.na(ld)] <- 0
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
ldGi <- ldweight_edges(gi, ld)
```

maxflow

*Maxflow algorithm***Description**

Run the maxflow algorithm.

**Usage**

```
maxflow(A, As, At)
```

**Arguments**

A	A sparse matrix with the connectivity.
As	A vector containing the edges to the source.
At	A vector containing the edges to the sink.

**Value**

A list with vector indicating if the feature was selected and the objective score.

mget\_gxg\_biogrid

*Memoised version of get\_gxg\_biogrid***Description**

Get all protein-protein interactions for an organism from BioGRID.

**Usage**

```
mget_gxg_biogrid(organism = 9606)
```

**Arguments**

organism	Tax ID of the studied organism. The default is 9606 (human).
----------	--------------------------------------------------------------

**Value**

A data.frame with two columns with pairs of interacting proteins.

**Examples**

```
# download dog interactions
## Not run: martini::get_gxg_biogrid(9615)
```

---

mget_gxg_string	<i>Memoised version of get_gxg_stringdb</i>
-----------------	---------------------------------------------

---

### Description

Get all protein-protein interactions for an organism from STRING. It uses a score cut-off of 400.

### Usage

```
mget_gxg_string(organism = 9606)
```

### Arguments

organism      Tax ID of the studied organism. The default is 9606 (human).

### Value

A data.frame with two columns with pairs of interacting proteins.

### Examples

```
# download frog interactions
## Not run: martini::get_gxg_string(8364)
```

---

mincut	<i>Run min-cut algorithm</i>
--------	------------------------------

---

### Description

Run min-cut algorithm

### Usage

```
mincut(gwas, net, covars, eta, lambda, score, sigmod, family, link)
```

### Value

A copy of the SnpMatrix\$map data.frame, with the following additions:

- c: contains the univariate association score for every single SNP.
- selected: logical vector indicating if the SNP was selected by SConES or not.
- module: integer with the number of the module the SNP belongs to.

---

mincut.cv

*Run the cross-validated min-cut algorithm*


---

**Description**

Run the cross-validated min-cut algorithm

**Usage**

```
mincut.cv(
  gwas,
  net,
  covars,
  etas,
  lambdas,
  criterion,
  score,
  sigmod,
  family,
  link,
  max_prop_snp
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.

---

mincut\_c

*Min-cut algorithm*


---

**Description**

Run the mincut algorithm.

**Usage**

```
mincut_c(c, eta, lambda, W)
```

**Arguments**

<code>c</code>	A vector with the association of each SNP with the phenotype.
<code>eta</code>	A numeric with the value of the eta parameter.
<code>lambda</code>	A numeric with the value of the eta parameter.
<code>W</code>	A sparse matrix with the connectivity.

**Value**

A list with vector indicating if the feature was selected and the objective score.

---

<code>minigwas</code>	<i>Description of the minigwas dataset.</i>
-----------------------	---------------------------------------------

---

**Description**

Small GWAS example.

**Format**

A list with 3 items:

**genotypes** Genotype and phenotype information.

**fam** Simulated network.

**map** Result of running `find_cones` with `gwas` and `net`.

**Examples**

```
data(minigwas)

# access different elements
minigwas[["genotypes"]]
minigwas[["map"]]
minigwas[["fam"]]
```

---

<code>minippi</code>	<i>PPIs for the minigwas dataset.</i>
----------------------	---------------------------------------

---

**Description**

`data.frame` describing pairs of proteins that interact for `minigwas`.

**Examples**

```
data(minippi)

head(minippi)
```



---

minisnpMapping	<i>Genes for the minigwas dataset.</i>
----------------	----------------------------------------

---

**Description**

data.frame that maps SNPs from minigwas to their gene.

**Examples**

```
data(minisnpMapping)
```

```
head(minisnpMapping)
```

---

organism_id2name	<i>Tax id to ensembl species name</i>
------------------	---------------------------------------

---

**Description**

Converts taxid to ensembl species name e.g. human databases are hsapiens\_\*

**Usage**

```
organism_id2name(id)
```

**Arguments**

organism      Tax ID of the studied organism. The default is 9606 (human).

---

permute_snpMatrix	<i>Permute samples</i>
-------------------	------------------------

---

**Description**

Compute a permutation of the samples of a snpMatrix object. Useful to make sure that the folds are not stratified by phenotype.

**Usage**

```
permute_snpMatrix(gwas)
```

**Arguments**

gwas            A SnpMatrix object with the GWAS information.

---

plot_ideogram	<i>Ideogram of SConES results.</i>
---------------	------------------------------------

---

**Description**

Create a circular ideogram of the a network results using the circlize package (Gu et al., 2014).

**Usage**

```
plot_ideogram(gwas, net, covars = data.frame(), genome = "hg19")
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
genome	Abbreviations of the genome to use: hg19 for human (default), mm10 for mouse, etc.

**Value**

A circular ideogram, including the manhattan plot, and the interactions between the selected SNPs.

**References**

Gu, Z., Gu, L., Eils, R., Schlesner, M., & Brors, B. (2014). circlize Implements and enhances circular visualization in R. *Bioinformatics* (Oxford, England), 30(19), 2811-2. <https://doi.org/10.1093/bioinformatics/btu393>

---

sanitize_map	<i>Check map</i>
--------------	------------------

---

**Description**

Check that map is a proper data.frame.

**Usage**

```
sanitize_map(gwas)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
------	-----------------------------------------------

---

sanitize\_snpMapping    *Check snpMapping*

---

**Description**

Check that snpMapping is a proper data.frame.

**Usage**

```
sanitize_snpMapping(snpMapping, col_genes)
```

**Arguments**

snpMapping	data.frame containing the correspondence between SNPs and genes.
col_genes	Length 2 character vector containing the colnames containing the SNP and the gene ids, respectively.

---

scones                      *Find connected explanatory SNPs*

---

**Description**

Finds the SNPs maximally associated with a phenotype while being connected in an underlying network.

**Usage**

```
scones(
  gwas,
  net,
  eta,
  lambda,
  covars = data.frame(),
  score = c("chi2", "glm", "r2"),
  family = c("binomial", "poisson", "gaussian", "gamma"),
  link = c("logit", "log", "identity", "inverse")
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
eta	Value of the eta parameter.
lambda	Value of the lambda parameter.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
score	Association score to measure association between genotype and phenotype. Possible values: chi2 (default), glm.

family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.

### Value

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

### References

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

### Examples

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
scones(minigwas, gi, 10, 1)
```

---

scones.cv

*Find connected explanatory SNPs.*

---

### Description

Finds the SNPs maximally associated with a phenotype while being connected in an underlying network. Select the hyperparameters by cross-validation.

### Usage

```
scones.cv(
  gwas,
  net,
  covars = data.frame(),
  score = c("chi2", "glm", "r2"),
  criterion = c("stability", "bic", "aic", "aicc", "global_clustering",
    "local_clustering"),
  etas = numeric(),
  lambdas = numeric(),
  family = c("binomial", "poisson", "gaussian", "gamma"),
  link = c("logit", "log", "identity", "inverse"),
  max_prop_snp = 0.5
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
score	Association score to measure association between genotype and phenotype. Possible values: chi2 (default), glm.
criterion	String with the function to measure the quality of a split.
etas	Numeric vector with the etas to explore in the grid search. If omitted, it's automatically created based on the association scores.
lambdas	Numeric vector with the lambdas to explore in the grid search. If omitted, it's automatically created based on the association scores.
family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.
max_prop_snp	Maximum proportion of SNPs accepted in the model (between 0 and 1). Larger solutions will be discarded.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**References**

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
scones.cv(minigwas, gi)
scones.cv(minigwas, gi, score = "glm")
```

---

scones.cv\_

*Find connected explanatory features*

---

**Description**

Finds the features maximally associated with a phenotype while being connected in an underlying network. Select the hyperparameters by cross-validation.

**Usage**

```
scones.cv_(X, y, featnames, net)
```

**Arguments**

X	n x d design matrix
y	Vector of length n with the outcomes
featnames	Vector of length d with the feature names
net	An igraph network that connects the SNPs.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**Examples**

```
X <- as(minigwas[['genotypes']], 'numeric')
X <- X + matrix(rnorm(2500, sd = 0.1), nrow(X), ncol(X))
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
scones.cv_(X, minigwas[['fam']]$affected, minigwas[['map']]$snp, gi)
```

---

scones\_

*Find connected explanatory features*

---

**Description**

Finds the features maximally associated with a phenotype while being connected in an underlying network.

**Usage**

```
scones_(X, y, featnames, net, eta, lambda)
```

**Arguments**

X	n x d design matrix
y	Vector of length n with the outcomes
featnames	Vector of length d with the feature names
net	An igraph network that connects the SNPs.
eta	Value of the eta parameter.
lambda	Value of the lambda parameter.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**Examples**

```
X <- as(minigwas[['genotypes']], 'numeric')
X <- X + matrix(rnorm(2500, sd = 0.1), nrow(X), ncol(X))
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
scones_(X, minigwas[['fam']]$affected, minigwas[['map']]$snp, gi, 10, 1)
```

---

score\_fold

*Score the solutions of a k-fold*

---

**Description**

Take the k-solutions for a combination of hyperparameters, and assign a score to it (the larger, the better).

**Usage**

```
score_fold(gwas, covars, net, selected, criterion, max_prop_snp)
```

**Arguments**

<code>gwas</code>	A <code>SnpMatrix</code> object with the GWAS information.
<code>covars</code>	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
<code>net</code>	An <code>igraph</code> network that connects the SNPs.
<code>criterion</code>	String with the function to measure the quality of a split.
<code>max_prop_snp</code>	Maximum proportion of SNPs accepted in the model (between 0 and 1). Larger solutions will be discarded.

---

search\_cones

*Find connected explanatory SNPs.*

---

**Description**

Finds the SNPs maximally associated with a phenotype while being connected in an underlying network (Azencott et al., 2013).

**Usage**

```
search_cones(
  gwas,
  net,
  encoding = "additive",
  sigmod = FALSE,
  covars = data.frame(),
  associationScore = c("chi2", "glm"),
  modelScore = c("stability", "bic", "aic", "aicc", "global_clustering",
    "local_clustering"),
  etas = numeric(),
  lambdas = numeric()
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
encoding	SNP encoding (unused argument).
sigmod	Boolean. If TRUE, use the Sigmod variant of SConES, meant to prioritize tightly connected clusters of SNPs.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
associationScore	Association score to measure association between genotype and phenotype.
modelScore	String with the function to measure the quality of a split.
etas	Numeric vector with the etas to explore in the grid search. If omitted, it's automatically created based on the association scores.
lambdas	Numeric vector with the lambdas to explore in the grid search. If omitted, it's automatically created based on the association scores.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**References**

Azencott, C. A., Grimm, D., Sugiyama, M., Kawahara, Y., & Borgwardt, K. M. (2013). Efficient network-guided multi-locus association mapping with graph cuts. *Bioinformatics*, 29(13), 171-179. <https://doi.org/10.1093/bioinformatics/btt238>

**Examples**

```
## Not run: gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
search_cones(minigwas, gi)
search_cones(minigwas, gi, encoding = "recessive")
search_cones(minigwas, gi, associationScore = "skat")
## End(Not run)
```



sigmod

*Find connected explanatory SNPs***Description**

Finds the SNPs maximally associated with a phenotype while being connected in an underlying network.

**Usage**

```
sigmod(
  gwas,
  net,
  eta,
  lambda,
  covars = data.frame(),
  score = c("chi2", "glm", "r2"),
  family = c("binomial", "poisson", "gaussian", "gamma"),
  link = c("logit", "log", "identity", "inverse")
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
eta	Value of the eta parameter.
lambda	Value of the lambda parameter.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
score	Association score to measure association between genotype and phenotype. Possible values: chi2 (default), glm.
family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**References**

Liu, Y., Brossard, M., Roqueiro, D., Margarithte-Jeannin, P., Sarnowski, C., Bouzigon, E., Demenais, F. (2017). SigMod: an exact and efficient method to identify a strongly interconnected disease-associated module in a gene network. *Bioinformatics*, 33(10), 1536–1544. <https://doi.org/10.1093/bioinformatics/btx004>

## Examples

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
sigmod(minigwas, gi, 10, 1)
```

---

sigmod.cv

*Find connected explanatory SNPs.*


---

## Description

Finds the SNPs maximally associated with a phenotype while being connected in an underlying network. Select the hyperparameters by cross-validation.

## Usage

```
sigmod.cv(
  gwas,
  net,
  covars = data.frame(),
  score = c("chi2", "glm", "r2"),
  criterion = c("stability", "bic", "aic", "aicc", "global_clustering",
    "local_clustering"),
  etas = numeric(),
  lambdas = numeric(),
  family = c("binomial", "poisson", "gaussian", "gamma"),
  link = c("logit", "log", "identity", "inverse"),
  max_prop_snp = 0.5
)
```

## Arguments

gwas	A SnpMatrix object with the GWAS information.
net	An igraph network that connects the SNPs.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
score	Association score to measure association between genotype and phenotype. Possible values: chi2 (default), glm.
criterion	String with the function to measure the quality of a split.
etas	Numeric vector with the etas to explore in the grid search. If omitted, it's automatically created based on the association scores.
lambdas	Numeric vector with the lambdas to explore in the grid search. If omitted, it's automatically created based on the association scores.
family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.
max_prop_snp	Maximum proportion of SNPs accepted in the model (between 0 and 1). Larger solutions will be discarded.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**References**

Liu, Y., Brossard, M., Roqueiro, D., Margaritte-Jeannin, P., Sarnowski, C., Bouzigon, E., Demenais, F. (2017). SigMod: an exact and efficient method to identify a strongly interconnected disease-associated module in a gene network. *Bioinformatics*, 33(10), 1536–1544. <https://doi.org/10.1093/bioinformatics/btx004>

**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
sigmod.cv(minigwas, gi)
sigmod.cv(minigwas, gi, score = "glm")
```

---

sigmod.cv\_

*Find connected explanatory features*

---

**Description**

Finds the features maximally associated with a phenotype while being connected in an underlying network. Select the hyperparameters by cross-validation.

**Usage**

```
sigmod.cv_(X, y, featnames, net)
```

**Arguments**

<code>X</code>	<code>n x d</code> design matrix
<code>y</code>	Vector of length <code>n</code> with the outcomes
<code>featnames</code>	Vector of length <code>d</code> with the feature names
<code>net</code>	An igraph network that connects the SNPs.

**Value**

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

**Examples**

```
X <- as(minigwas[['genotypes']], 'numeric')
X <- X + matrix(rnorm(2500, sd = 0.1), nrow(X), ncol(X))
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
sigmod.cv_(X, minigwas[['fam']]$affected, minigwas[['map']]$snp, gi)
```

---

sigmod\_ *Find connected explanatory features*

---

### Description

Finds the features maximally associated with a phenotype while being connected in an underlying network.

### Usage

```
sigmod_(X, y, featnames, net, eta, lambda)
```

### Arguments

X	n x d design matrix
y	Vector of length n with the outcomes
featnames	Vector of length d with the feature names
net	An igraph network that connects the SNPs.
eta	Value of the eta parameter.
lambda	Value of the lambda parameter.

### Value

A copy of the `SnpMatrix$map` data.frame, with the following additions:

- `c`: contains the univariate association score for every single SNP.
- `selected`: logical vector indicating if the SNP was selected by SConES or not.
- `module`: integer with the number of the module the SNP belongs to.

### Examples

```
X <- as(minigwas[['genotypes']], 'numeric')
X <- X + matrix(rnorm(2500, sd = 0.1), nrow(X), ncol(X))
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
sigmod_(X, minigwas[['fam']]$affected, minigwas[['map']]$snp, gi, 10, 1)
```

---

simulate\_causal\_snps *Simulate causal SNPs*

---

### Description

Selects randomly interconnected genes as causal, then selects a proportion of them as causal.

### Usage

```
simulate_causal_snps(net, ngenes = 20, pcausal = 1)
```

**Arguments**

net	An igraph gene-interaction (GI) network that connects the SNPs.
ngenes	Number of causal genes.
pcausal	Number between 0 and 1, proportion of the SNPs in causal genes that are causal themselves.

**Value**

A vector with the ids of the simulated causal SNPs.

**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
simulate_causal_snps(gi, ngenes=2)
```

---

simulate_phenotype	<i>Simulate phenotype</i>
--------------------	---------------------------

---

**Description**

Simulates a phenotype from a GWAS experiment and a specified set of causal SNPs. If the data is qualitative, only controls are used.

**Usage**

```
simulate_phenotype(
  gwas,
  snps,
  h2,
  model = "additive",
  effectSize = rnorm(length(snps)),
  qualitative = FALSE,
  ncases,
  ncontrols,
  prevalence
)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
snps	Character vector with the SNP ids of the causal SNPs. Must match SNPs in <code>gwas[["map"]][["snp.name"]]</code> .
h2	Heritability of the phenotype (between 0 and 1).
model	String specifying the genetic model under the phenotype. Accepted values: "additive".
effectSize	Numeric vector with the same length as the number of causal SNPs. It indicates the effect size of each of the SNPs; if absent, they are sampled from a normal distribution.
qualitative	Bool indicating if the phenotype is qualitative or not (quantitative).

ncases	Integer specifying the number of cases to simulate in a qualitative phenotype. Required if qualitative = TRUE.
ncontrols	Integer specifying the number of controls to simulate in a qualitative phenotype. Required if qualitative = TRUE.
prevalence	Value between 0 and 1 specifying the population prevalence of the disease. Note that ncases cannot be greater than prevalence * number of samples. Required if qualitative = TRUE.

**Value**

A copy of the GWAS experiment with the new phenotypes in `gwas[["fam"]][["affected"]]`.

**References**

Inspired from GCTA simulation tool: <http://cnsgenomics.com/software/gcta/Simu.html>.

**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
causal <- simulate_causal_snps(gi, ngenes = 2)
simulate_phenotype(minigwas, causal, h2 = 1)
```

---

snp2ensembl	<i>Map SNPs to Ensembl genes.</i>
-------------	-----------------------------------

---

**Description**

Maps SNPs from a GWAS experiment to genes.

**Usage**

```
snp2ensembl(gwas, organism = 9606, flank = 0)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
organism	Tax ID of the studied organism. The default is 9606 (human).
flank	A number with the flanking regions around genes to be considered part of the gene i.e. SNPs mapped to them will be considered mapped to the gene.

**Value**

A data.frame with two columns: one for the SNP and another for the gene it has been mapped to.

---

snp_test	<i>Calculate genotype-phenotype associations</i>
----------	--------------------------------------------------

---

**Description**

Calculate the association between genotypes and a phenotype, adjusting by covariates.

**Usage**

```
snp_test(gwas, covars, score, family, link)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
covars	A data frame with the covariates. It must contain a column 'sample' containing the sample IDs, and an additional columns for each covariate.
score	Association score to measure association between genotype and phenotype. Possible values: chi2 (default), glm.
family	A string defining the generalized linear model family. This should match one of "binomial", "poisson", "gaussian" or "gamma". See <a href="#">snp.rhs.tests</a> for details.
link	A string defining the link function for the GLM. This should match one of "logit", "log", "identity" or "inverse". See <a href="#">snp.rhs.tests</a> for details.

**Value**

A named vector with the association scores.

---

subnet	<i>Subgraph of vertices with an attribute</i>
--------	-----------------------------------------------

---

**Description**

Returns a subgraph matching some condition.

**Usage**

```
subnet(net, attr, values, affirmative = TRUE)
```

**Arguments**

net	An igraph network.
attr	An attribute of the vertices.
values	Possible values of attr.
affirmative	Logical. States if a condition must be its affirmation (e.g. all nodes with gene name "X"), or its negation (all nodes not with gene name "X").

**Value**

A subgraph containing only the vertices with attribute equal to any of the values in values.

**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
martini:::subnet(gi, "gene", "A")
martini:::subnet(gi, "name", c("1A1", "1A3"))
```

---

subset_snpMatrix	<i>Subsample snpMatrix</i>
------------------	----------------------------

---

**Description**

Compute a permutation of the samples of a snpMatrix object. Useful to make sure that the folds are not stratified by phenotype.

**Usage**

```
subset_snpMatrix(gwas, samples)
```

**Arguments**

gwas	A SnpMatrix object with the GWAS information.
samples	Vector (logical or numeric) containing the samples to select.

---

subvert	<i>Vertices with an attribute</i>
---------	-----------------------------------

---

**Description**

Returns the nodes matching some condition.

**Usage**

```
subvert(net, attr, values, affirmative = TRUE)
```

**Arguments**

net	An igraph network.
attr	An attribute of the vertices.
values	Possible values of attr
affirmative	Logical. States if a condition must be its affirmation (e.g. all nodes with gene name "X"), or its negation (all nodes not with gene name "X").

**Value**

The vertices with attribute equal to any of the values in values.



**Examples**

```
gi <- get_GI_network(minigwas, snpMapping = minisnpMapping, ppi = minippi)
martini:::subvert(gi, "gene", "A")
martini:::subvert(gi, "name", c("1A1", "1A3"))
```

---

`wrap_Xy`*Make pseudo SnpMatrix object*

---

**Description**

Wrap design matrix and outcome vector into a pseudo SnpMatrix object.

**Usage**

```
wrap_Xy(X, y, featnames, net)
```

**Arguments**

<code>X</code>	n x d design matrix
<code>y</code>	Vector of length n with the outcomes
<code>featnames</code>	Vector of length d with the feature names
<code>net</code>	An igraph network that connects the SNPs.

# Index

## \* internal

- arrange\_covars, 3
- calculateE, 3
- calculateG, 4
- check\_installed, 4
- connect\_biomart, 5
- get\_adjacency, 5
- get\_grid, 8
- get\_gxg, 9
- get\_gxg\_biogrid, 9
- get\_gxg\_string, 10
- get\_snp\_modules, 10
- group\_snps, 11
- is\_coherent, 12
- mget\_gxg\_biogrid, 13
- mget\_gxg\_string, 14
- mincut, 14
- mincut.cv, 15
- organism\_id2name, 17
- permute\_snpMatrix, 17
- sanitize\_map, 18
- sanitize\_snpMapping, 19
- score\_fold, 23
- snp2ensembl, 30
- snp\_test, 31
- subnet, 31
- subset\_snpMatrix, 32

arrange\_covars, 3

calculateE, 3  
calculateG, 4  
check\_installed, 4  
connect\_biomart, 5

get\_adjacency, 5  
get\_GI\_network, 6  
get\_GM\_network, 7  
get\_grid, 8  
get\_GS\_network, 8  
get\_gxg, 9  
get\_gxg\_biogrid, 9  
get\_gxg\_string, 10  
get\_snp\_modules, 10

GM, 6

group\_snps, 11  
GS, 7  
gwas2bed, 11

is\_coherent, 12

ld, 12  
ldweight\_edges, 12

maxflow, 13  
mget\_gxg\_biogrid, 13  
mget\_gxg\_string, 14  
mincut, 14  
mincut.cv, 15  
mincut\_c, 15  
minigwas, 16  
minippi, 16  
minisnpMapping, 17

organism\_id2name, 17

permute\_snpMatrix, 17  
plot\_ideogram, 18

sanitize\_map, 18  
sanitize\_snpMapping, 19  
scones, 19

scones.cv, 20  
scones.cv\_, 21  
scones\_, 22  
score\_fold, 23  
search\_cones, 23

sigmoid, 25  
sigmoid.cv, 26  
sigmoid.cv\_, 27  
sigmoid\_, 28

simulate\_causal\_snps, 28  
simulate\_phenotype, 29  
snp.rhs.tests, 15, 20, 21, 25, 26, 31  
snp2ensembl, 30  
snp\_test, 31  
subnet, 31  
subset\_snpMatrix, 32  
subvert, 32

wrap\_Xy, [33](#)