

Package ‘prebs’

January 19, 2021

Title Probe region expression estimation for RNA-seq data for improved microarray comparability

Description The prebs package aims at making RNA-sequencing (RNA-seq) data more comparable to microarray data. The comparability is achieved by summarizing sequencing-based expressions of probe regions using a modified version of RMA algorithm. The pipeline takes mapped reads in BAM format as an input and produces either gene expressions or original microarray probe set expressions as an output.

Version 1.30.0

Author Karolis Uziela and Antti Honkela

Maintainer Karolis Uziela <karolis.uziela@scilifelab.se>

Depends R (>= 2.14.0), GenomicAlignments, affy, RPA

Imports parallel, methods, stats, GenomicRanges (>= 1.13.3), IRanges, Biobase, GenomeInfoDb, S4Vectors

Suggests prebsdata, hgu133plus2cdf, hgu133plus2probe

License Artistic-2.0

Collate 'PREBS.R'

biocViews ImmunoOncology, Microarray, RNASeq, Sequencing, GeneExpression, Preprocessing

git_url <https://git.bioconductor.org/packages/prebs>

git_branch RELEASE_3_12

git_last_commit 24d9c3a

git_last_commit_date 2020-10-27

Date/Publication 2021-01-18

R topics documented:

calc_prebs	2
prebs	3

Index	5
--------------	----------

calc_prebs	<i>Calculate PREBS values</i>
------------	-------------------------------

Description

calc_prebs calculates PREBS values for given set of BAM files.

Usage

```
calc_prebs(bam_files, probe_mapping_file, cdf_name = NULL, cluster = NULL,
           output_eset = TRUE, paired_ended_reads = FALSE, ignore_strand = TRUE,
           sum.method = "rpa")
```

Arguments

bam_files	A vector containing .bam files.
probe_mapping_file	A file containing probe mappings in the genome.
cdf_name	A name of CDF package to use in RMA algorithm. If cdf_name=NULL, the package name is inferred from the name of probe_mapping_file ("HGU133Plus2_Hs_ENSG_mapping" -> "hgu133plus2hsensgcdf")
cluster	A cluster object created using "makeCluster" function from "parallel" package. If cluster=NULL, no parallelization is used.
output_eset	If set to TRUE, the output of calc_prebs will be ExpressionSet object. Otherwise, the output will be a data frame.
paired_ended_reads	Set it to TRUE if your data contains paired-ended reads. Otherwise, the two read mates will be treated as independent units.
ignore_strand	If set to TRUE, then the strand is ignored while counting read overlaps with probe regions. If you use strand-specific RNA-seq protocol, set to FALSE, otherwise set it to TRUE.
sum.method	Microarray summarization method to be used. Can be either rpa or rma. The default mode is rpa.

Details

calc_prebs is the main function of prebs package that implements the whole pipeline. The function takes mapped reads in BAM format and probe sequence mappings as an input.

calc_prebs can run in two modes: rpa and rma. RMA is the classical microarray summarization algorithm developed by R. A. Irizarry et al. (2003), while RPA is a newer algorithm that was developed by L. Lahti et al. (2011). The default mode is rpa. NOTE: before prebs version 1.7.1 only RMA mode was available.

The output format depends on output_eset option. If output_eset=TRUE then calc_prebs returns ExpressionSet object (ExpressionSet object is defined in affy package). Otherwise, it returns a data frame containing PREBS values.

For running calc_prebs with custom CDF, the custom CDF package has to be downloaded and installed from Custom CDF website: <http://brainarray.mbni.med.umich.edu/CustomCDF>

For running calc_prebs with manufacturer's CDF, the manufacturer's CDF package can be installed from Bioconductor, for example: `BiocManager::install("GenomicRanges"); BiocManager::install("hgu133plus2c`

For a detailed input specification, please refer to the prebs vignette.

Value

ExpressionSet object or a data frame containing PREBS values

Examples

```

if (require(prebsdata)) {
  # Get full paths to data files in \code{prebsdata} package
  bam_file1 <- system.file(file.path("sample_bam_files", "input1.bam"), package="prebsdata")
  bam_file2 <- system.file(file.path("sample_bam_files", "input2.bam"), package="prebsdata")
  bam_files <- c(bam_file1, bam_file2)
  custom_cdf_mapping1 <- system.file(file.path("custom-cdf", "HGU133Plus2_Hs_ENSG_mapping.txt"),
                                     package="prebsdata")
  custom_cdf_mapping2 <- system.file(file.path("custom-cdf", "HGU133A2_Hs_ENSG_mapping.txt"),
                                     package="prebsdata")
  manufacturer_cdf_mapping <- system.file(file.path("manufacturer-cdf", "HGU133Plus2_mapping.txt"),
                                          package="prebsdata")

  if (interactive()) {
    # Run PREBS using custom CDF without parallelization ("rpa" mode)
    prebs_values <- calc_prebs(bam_files, custom_cdf_mapping1)
    head(exprs(prebs_values))

    # Run PREBS using custom CDF without parallelization ("rma" mode)
    prebs_values <- calc_prebs(bam_files, custom_cdf_mapping1, sum.method="rma")
    head(exprs(prebs_values))

    # Run PREBS using custom CDF with parallelization
    library(parallel)
    N_CORES = 2
    CLUSTER <- makeCluster(N_CORES)
    prebs_values <- calc_prebs(bam_files, custom_cdf_mapping1, cluster=CLUSTER)
    stopCluster(CLUSTER)

    # Run PREBS using another custom CDF
    prebs_values <- calc_prebs(bam_files, custom_cdf_mapping2)

    # Run PREBS and return data frame instead of ExpressionSet object
    prebs_values <- calc_prebs(bam_files, custom_cdf_mapping1, output_eset=FALSE)
    head(prebs_values)
  }

  # Run PREBS using Manufacturer's CDF (outputs probe set expressions)
  prebs_values <- calc_prebs(bam_files, manufacturer_cdf_mapping)
  head(exprs(prebs_values))

  # Same as above, but state CDF package name explicitly
  prebs_values <- calc_prebs(bam_files, manufacturer_cdf_mapping, cdf_name="hgu133plus2cdf")
}

```

Description

The *prebs* package aims at making RNA-sequencing (RNA-seq) data more comparable to microarray data. The comparability is achieved by summarizing sequencing-based expressions of probe regions using standard microarray summarization algorithms (RPA or RMA). The pipeline takes mapped reads in BAM format as an input and produces either gene expressions or original microarray probe set expressions as an output.

Details

The package has only one public function: `calc_prebs`. Type `help(calc_prebs)` for more information on the usage.

Index

[calc_prebs](#), 2

[prebs](#), 3

[prebs-package \(prebs\)](#), 3