

Package ‘crossmeta’

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Title Cross Platform Meta-Analysis of Microarray Data

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Author Alex Pickering

Maintainer Alex Pickering <alexvpickering@gmail.com>

Description Implements cross-platform and cross-species meta-analyses of Affymetrix, Illumina, and Agilent microarray data. This package automates common tasks such as downloading, normalizing, and annotating raw GEO data. The user then selects control and treatment samples in order to perform differential expression analyses for all comparisons. After analysing each contrast separately, the user can select tissue sources for each contrast and specify any tissue sources that should be grouped for the subsequent meta-analyses.

Depends R (>= 4.0)

SystemRequirements libxml2: libxml2-dev (deb), libxml2-devel (rpm)
libcurl: libcurl4-openssl-dev (deb), libcurl-devel (rpm)
openssl: libssl-dev (deb), openssl-devel (rpm), libssl_dev
(csw), openssl@1.1 (brew)

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Encoding UTF-8

LazyData TRUE

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BugReports <https://github.com/alexvpickering/crossmeta/issues>

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Contents

addContrastInput	3
add_adjusted	4
add_sources	4
add_vsd	5
bulkAnnot	6
bulkAnnotInput	6
bulkForm	6
bulkFormInput	7
bulkPage	7
bulkPageUI	7
bulkTable	8
bulkTableOutput	8
ch2_subset	8
clean_y	9
delContrastsInput	9
diff_expr	10
es_meta	11
exprs.MA	13
filter_genes	13
fit_ebayes	14
fit_lm	15
fix_illum_headers	15
format_dl_annot	16
format_up_annot	16
get_ch2_mod	16
get_group_levels	17
get_palette	17
get_raw	18
get_sva_mods	18
get_top_table	19

get_vsd	20
gs.names	20
gslist	21
ilmn.nnum	21
iqr_replicates	22
is_invertible	22
load_agil_plat	23
load_diff	23
load_illum_plat	24
load_plat	24
load_raw	25
makeExampleCountsEset	26
match_prev_eset	27
open_raw_illum	27
phenoData.ch2	28
prefix_illum_headers	28
query_ref	29
remove_autonamed	29
run_limma	30
run_limma_setup	31
run_lmfit	31
run_select_contrasts	32
run_sva	32
setup_prev	33
symbol_annot	34
to_eset	35
to_ma	35
validate_up_annot	36
which_max_iqr	36
xls_to_txt	37
Index	38

addContrastInput	<i>Add contrast input</i>
------------------	---------------------------

Description

Add contrast input

Usage

addContrastInput(id)

add_adjusted	<i>Add expression data adjusted for pairs/surrogate variables</i>
--------------	---

Description

Add expression data adjusted for pairs/surrogate variables

Usage

```
add_adjusted(eset, svobj = list(sv = NULL), numsv = 0)
```

Arguments

eset	ExpressionSet
svobj	surrogate variable object
numsv	Number of surrogate variables to adjust for

Value

eset with adjusted element added

add_sources	<i>Add sample source information for meta-analysis.</i>
-------------	---

Description

User selects a tissue source for each contrast and indicates any sources that should be paired. This step is required if you would like to perform source-specific effect-size/pathway meta-analyses.

Usage

```
add_sources(diff_exprs, data_dir = getwd(), postfix = NULL)
```

Arguments

diff_exprs	Previous result of <code>diff_expr</code> , which can be reloaded using <code>load_diff</code> .
data_dir	String specifying directory of GSE folders.
postfix	Optional string to append to saved results. Useful if need to run multiple meta-analyses on the same series but with different contrasts.

Details

The **Sources** tab is used to add a source for each contrast. To do so: click the relevant contrast rows, search for a source in the *Sample source* dropdown box, and then click the *Add* button.

The **Pairs** tab is used to indicate sources that should be paired (treated as the same source for subsequent effect-size and pathway meta-analyses). To do so: select at least two sources from the *Paired sources* dropdown box, and then click the *Add* button.

For each GSE, analysis results with added sources/pairs are saved in the corresponding GSE folder (in `data_dir`) that was created by `get_raw`.

Value

Same as `diff_expr` with added slots for each GSE in `diff_exprs`:

<code>sources</code>	Named vector specifying selected sample source for each contrast. Vector names identify the contrast.
<code>pairs</code>	List of character vectors indicating tissue sources that should be treated as the same source for subsequent effect-size and pathway meta-analyses.

Examples

```
library(lydata)

# load result of previous call to diff_expr:
data_dir <- system.file("extdata", package = "lydata")
gse_names <- c("GSE9601", "GSE34817")
anals <- load_diff(gse_names, data_dir)

# run shiny GUI to add tissue sources
# anals <- add_sources(anals, data_dir)
```

<code>add_vsd</code>	<i>Add VST normalized assay data element to expression set</i>
----------------------	--

Description

For microarray datasets duplicates `exprs` slot into `vsd` slot.

Usage

```
add_vsd(eset, rna_seq = TRUE)
```

Arguments

<code>eset</code>	ExpressionSet with group column in <code>pData(eset)</code>
<code>rna_seq</code>	Is this an RNA-seq eset? Default is TRUE.

Value

eset with 'vsd' assayDataElement added.

bulkAnnot	<i>Logic for downloading and uploading bulk annotation</i>
-----------	--

Description

Logic for downloading and uploading bulk annotation

Usage

```
bulkAnnot(input, output, session, dataset_name, pdata)
```

bulkAnnotInput	<i>UI for Bulk Data annotation upload/download</i>
----------------	--

Description

UI for Bulk Data annotation upload/download

Usage

```
bulkAnnotInput(id)
```

bulkForm	<i>Logic for Bulk Data form</i>
----------	---------------------------------

Description

Logic for Bulk Data form

Usage

```
bulkForm(input, output, session, pdata, prev)
```

bulkFormInput	<i>Input form for Bulk Data page</i>
---------------	--------------------------------------

Description

Input form for Bulk Data page

Usage

```
bulkFormInput(id)
```

bulkPage	<i>Logic for Select Contrasts Interface</i>
----------	---

Description

Logic for Select Contrasts Interface

Usage

```
bulkPage(input, output, session, eset, gse_name, prev)
```

Arguments

input, output, session	shiny module boilerplate
eset	ExpressionSet
gse_name	GEO accession for the series.
prev	Previous result of diff_expr. Used to allow rechecking previous selections.

bulkPageUI	<i>UI for Select Contrasts Interface</i>
------------	--

Description

UI for Select Contrasts Interface

Usage

```
bulkPageUI(id)
```

Arguments

id	The id string to be namespaced.
----	---------------------------------

bulkTable	<i>Logic for pdata table</i>
-----------	------------------------------

Description

Logic for pdata table

Usage

```
bulkTable(input, output, session, eset, prev, up_annot)
```

bulkTableOuput	<i>Tables for datasets page</i>
----------------	---------------------------------

Description

Tables for datasets page

Usage

```
bulkTableOuput(id)
```

ch2_subset	<i>Subset for Paired Two-Channel ExpressionSet</i>
------------	--

Description

Two-channel esets use intraspotCorrelation and lmscFit so can't use duplicateCorrelation. If not using one channel in contrasts (e.g. because all reference RNA) and have paired design, better to treat as single channel so that can use duplicateCorrelation.

Usage

```
ch2_subset(eset, prev_anal)
```

Arguments

eset	Annotated ExpressionSet. Created by load_raw.
prev_anal	One item (for eset) from previous result of diff_expr.

Value

ExpressionSet. If two-channel, paired and one channel not used will subset to used channel.

clean_y	<i>Adjusts expression data for surrogate variables.</i>
---------	---

Description

Factors out effect of surrogate variables discovered during surrogate variable analysis.

Usage

```
clean_y(y, mod, mod.clean)
```

Arguments

y	Expression data of eset.
mod	Full model matrix supplied to sva.
mod.clean	Model matrix with factors to clean.

Value

Expression data with effects of svcs removed.

delContrastsInput	<i>Delete contrasts input</i>
-------------------	-------------------------------

Description

Delete contrasts input

Usage

```
delContrastsInput(id)
```

diff_expr

*Differential expression analysis of esets.***Description**

After selecting control and test samples for each contrast, surrogate variable analysis ([sva](#)) and differential expression analysis is performed.

Usage

```
diff_expr(
  esets,
  data_dir = getwd(),
  annot = "SYMBOL",
  prev_anals = list(NULL),
  svanal = TRUE,
  recheck = FALSE,
  postfix = NULL,
  port = 3838
)
```

Arguments

esets	List of annotated esets. Created by load_raw .
data_dir	String specifying directory of GSE folders.
annot	String, column name in fData common to all esets. For duplicated values in this column, the row with the highest interquartile range across selected samples will be kept. If meta-analysis will follow, appropriate values are "SYMBOL" (default - for gene level analysis) or, if all esets are from the same platform, "PROBE" (for probe level analysis).
prev_anals	Previous result of diff_expr , which can be reloaded using load_diff . If present, previous selections, names, and pairs will be reused.
svanal	Use surrogate variable analysis? Default is TRUE.
recheck	Would you like to recheck previous group/contrast annotations? Requires <code>prev_anals</code> . Default is FALSE.
postfix	Optional string to append to saved results. Useful if need to run multiple meta-analyses on the same series but with different contrasts.
port	See runApp() .

Details

Click the Download icon and fill in the *Group name* column and optionally the *Pairs* column. Then save and upload the filled in metadata csv. After doing so, select a test and control group to compare and click the + icon to add the contrast. Repeat previous step to add additional contrasts.

After control and test samples have been added for all contrasts that you wish to include, click the *Done* button. Repeat for all GSEs.

Paired samples (e.g. the same subject before and after treatment) can be specified by filling out the *Pairs column* before uploading the metadata.

For each GSE, analysis results are saved in the corresponding GSE folder in `data_dir` that was created by `get_raw`. If analyses needs to be repeated, previous results can be reloaded with `load_diff` and supplied to the `prev_anals` parameter. In this case, previous selections, names, and pairs will be reused.

Value

List of named lists, one for each GSE. Each named list contains:

<code>pdata</code>	data.frame with phenotype data for selected samples. Columns <code>treatment</code> ('ctrl' or 'test'), <code>group</code> , and <code>pair</code> are added based on user selections.
<code>top_tables</code>	List with results of <code>topTable</code> call (one per contrast). These results account for the effects of nuisance variables discovered by surrogate variable analysis.
<code>ebayes_sv</code>	Results of call to <code>eBayes</code> with surrogate variables included in the model matrix.
<code>annot</code>	Value of <code>annot</code> variable.

Examples

```
library(lydata)

# location of raw data
data_dir <- system.file("extdata", package = "lydata")

# gather GSE names
gse_names <- c("GSE9601", "GSE15069", "GSE50841", "GSE34817", "GSE29689")

# load first eset
esets <- load_raw(gse_names[1], data_dir)

# run analysis (opens GUI)
# anal_old <- diff_expr(esets, data_dir)

# re-run analysis on first eset
prev <- load_diff(gse_names[1], data_dir)
anals <- diff_expr(esets[1], data_dir, prev_anals = prev)
```

es_meta

Effect size combination meta analysis.

Description

Performs effect-size meta-analyses across all studies and seperately for each tissue source.

Usage

```
es_meta(diff_exprs, cutoff = 0.3, by_source = FALSE)
```

Arguments

diff_exprs	Previous result of <code>diff_expr</code> , which can be reloaded using <code>load_diff</code> .
cutoff	Minimum fraction of contrasts that must have measured each gene. Between 0 and 1.
by_source	Should separate meta-analyses be performed for each tissue source added with <code>add_sources</code> ?

Details

Builds on `zScores` function from GeneMeta by allowing for genes that were not measured in all studies. This implementation also uses moderated unbiased effect sizes calculated by `effectsize` from metaMA and determines false discovery rates using `fdrtool`.

Value

A list of named lists, one for each tissue source. Each list contains two named data.frames. The first, `filt`, has all the columns below for genes present in cutoff or more fraction of contrasts. The second, `raw`, has only `dprime` and `vardprime` columns, but for all genes (NAs for genes not measured by a given contrast).

dprime	Unbiased effect sizes (one column per contrast).
vardprime	Variances of unbiased effect sizes (one column per contrast).
mu	Overall mean effect sizes.
var	Variances of overall mean effect sizes.
z	Overall z score = $\mu / \sqrt{\text{var}}$.
fdr	False discovery rates calculated from column z using <code>fdrtool</code> .
pval	p-values calculated from column z using <code>fdrtool</code> .

Examples

```
library(lydata)

# location of data
data_dir <- system.file("extdata", package = "lydata")

# gather GSE names
gse_names <- c("GSE9601", "GSE15069", "GSE50841", "GSE34817", "GSE29689")

# load previous analysis
anals <- load_diff(gse_names, data_dir)

# add tissue sources to perform separate meta-analyses for each source (optional)
# anals <- add_sources(anals, data_dir)
```

```
# perform meta-analysis
es <- es_meta(anals, by_source = TRUE)
```

 exprs.MA

Extract Log-Expression Matrix from MAList

Description

Converts M and A-values to log-expression values. The output matrix will have two columns for each array, in the order all red then all green. Adapted from [plotDensities.MAList](#) instead of [exprs.MA](#) so that order is same as [phenoData.ch2](#).

Usage

```
exprs.MA(MA)
```

Arguments

MA an MAList object.

Value

A numeric matrix with twice the columns of the input.

 filter_genes

Filter genes in RNA-seq ExpressionSet

Description

Uses [filterByExpr](#) to filter based on 'counts' assay or 'exprs' assay if 'counts' isn't available (for ARCHS4 data).

Usage

```
filter_genes(eset)
```

Arguments

eset ExpressionSet with 'counts' assayDataElement and group column in pData

Value

filtered eset

See Also[filterByExpr](#)**Examples**

```
# example ExpressionSet
eset <- makeExampleCountsEset()
eset <- filter_genes(eset)
```

`fit_ebayes`*Fit ebayes model*

Description

Fit ebayes model

Usage

```
fit_ebayes(
  lm_fit,
  contrasts,
  robust = TRUE,
  trend = FALSE,
  allow.no.resid = FALSE
)
```

Arguments

<code>lm_fit</code>	Result of call to run_limma
<code>contrasts</code>	Character vector of contrasts to fit.
<code>robust</code>	logical, should the estimation of <code>df.prior</code> and <code>var.prior</code> be robustified against outlier sample variances?
<code>trend</code>	logical, should an intensity-dependent trend be allowed for the prior variance? If FALSE then the prior variance is constant. Alternatively, <code>trend</code> can be a row-wise numeric vector, which will be used as the covariate for the prior variance.
<code>allow.no.resid</code>	Allow no residual degrees of freedom? if TRUE and the fit contrast matrix has no residual degrees of freedom, eBayes fit is skipped and the result of contrasts.fit is returned.

Valueresult of [eBayes](#)

fit_lm	<i>Run limma analysis.</i>
--------	----------------------------

Description

Runs limma differential expression analysis on all contrasts selected by `add_contrast`. Analysis performed with and without surrogate variables discovered by `diff_setup`. Also prints MDS plot and saves results.

Usage

```
fit_lm(eset, svobj = list(sv = NULL), numsv = 0, rna_seq = TRUE)
```

Arguments

eset	Annotated eset created by <code>load_raw</code> . Replicate features and non-selected samples removed by <code>iqr_replicates</code> .
svobj	Surrogate variable analysis results. Returned from <code>run_sva</code> .
numsv	Number of surrogate variables to model.
rna_seq	Is this an RNA-seq eset? Default is TRUE.

Value

list with slots: * fit Result of `lmFit`. * mod model matrix used for fit.

fix_illum_headers	<i>Attempts to fix Illumina raw data header</i>
-------------------	---

Description

Reads raw data files and tries to fix them up so that they can be loaded by `read.ilmn`.

Usage

```
fix_illum_headers(elist_paths, eset = NULL)
```

Arguments

elist_paths	Path to Illumina raw data files. Usually contain patterns: <code>non_normalized.txt</code> , <code>raw.txt</code> , or <code>_supplementary_.txt</code>
eset	ExpressionSet from <code>getGEO</code> .

Value

Character vector for annotation argument to `read.ilmn`. Fixed raw data files are saved with file-name ending in `_fixed.txt`

format_dl_annot	<i>Format downloaded annotation</i>
-----------------	-------------------------------------

Description

Format downloaded annotation

Usage

```
format_dl_annot(annot)
```

format_up_annot	<i>Format uploaded annotation</i>
-----------------	-----------------------------------

Description

Format uploaded annotation

Usage

```
format_up_annot(up, ref)
```

get_ch2_mod	<i>Get design matrix for two-channel array</i>
-------------	--

Description

Get design matrix for two-channel array

Usage

```
get_ch2_mod(eset)
```

Arguments

eset	ExpressionSet with colnames that end in '_red' and '_green' indicating channel and eset\$group indicating group membership.
------	---

Value

model matrix for use by [inraspotCorrelation](#) and [lmscFit](#)

`get_group_levels` *Get group levels for bulk data plots*

Description

Get group levels for bulk data plots

Usage

```
get_group_levels(pdata)
```

Arguments

`pdata` Data.frame of phenotype data

`get_palette` *Get a Palette to Distinguish Groups*

Description

Get a Palette to Distinguish Groups

Usage

```
get_palette(levs, dark = FALSE, with_all = FALSE)
```

Arguments

`levs` Character vector of levels to get colour palette for.

Value

Character vector with colour codes of length(`levs`).

get_raw	<i>Download and unpack microarray supplementary files from GEO.</i>
---------	---

Description

Downloads and unpacks microarray supplementary files from GEO. Files are stored in the supplied data directory under the GSE name.

Usage

```
get_raw(gse_names, data_dir = getwd())
```

Arguments

gse_names	Character vector of GSE names to download.
data_dir	String specifying directory for GSE folders.

Value

NULL (for download/unpack only).

See Also

[load_raw](#).

Examples

```
get_raw("GSE41845")
```

get_sva_mods	<i>Get model matrices for surrogate variable analysis</i>
--------------	---

Description

Used by add_adjusted to create model matrix with surrogate variables.

Usage

```
get_sva_mods(pdata)
```

Arguments

pdata	data.frame of phenotype data with column 'group' and 'pair' (optional).
-------	---

Value

List with model matrix(mod) and null model matrix (mod0) used for sva.

get_top_table	<i>Get top table</i>
---------------	----------------------

Description

Get top table

Usage

```
get_top_table(  
  lm_fit,  
  groups = c("test", "ctrl"),  
  with.es = TRUE,  
  robust = FALSE,  
  trend = FALSE,  
  allow.no.resid = FALSE  
)
```

Arguments

lm_fit	Result of run_limma
groups	Test and Control group as strings.
with.es	Add 'dprime' and 'vardprime' from effectsize ? Default is TRUE.
robust	logical, should the estimation of <code>df.prior</code> and <code>var.prior</code> be robustified against outlier sample variances?
trend	logical, should an intensity-dependent trend be allowed for the prior variance? If FALSE then the prior variance is constant. Alternatively, <code>trend</code> can be a row-wise numeric vector, which will be used as the covariate for the prior variance.
allow.no.resid	Allow no residual degrees of freedom? if TRUE and the fit contrast matrix has no residual degrees of freedom, eBayes fit is skipped and the result of contrasts.fit is returned.

Value

result of [toptable](#)

`get_vsd`*Get variance stabilized data for exploratory data analysis*

Description

Get variance stabilized data for exploratory data analysis

Usage

```
get_vsd(eset)
```

Arguments

`eset` ExpressionSet loaded with [load_raw](#).

Value

matrix with variance stabilized expression data.

`gs.names`*Map between KEGG pathway numbers and names.*

Description

Used to map human KEGG pathway numbers to names. Updated Feb 2017.

Usage

```
data(gs.names)
```

Format

An object of class character of length 310.

Value

A named character vector of human KEGG pathway names. Names of vector are KEGG pathway numbers.

gslis	<i>KEGG human pathway genes.</i>
-------	----------------------------------

Description

Genes for human KEGG pathways. Updated Feb 2017.

Usage

```
data(gslis)
```

Format

An object of class `list` of length 310.

Value

A named list with entrez ids of genes for human KEGG pathways. List names are KEGG pathway numbers.

ilmn.nnum	<i>Count numeric columns in raw Illumina data files</i>
-----------	---

Description

Excludes probe ID cols

Usage

```
ilmn.nnum(elist_paths)
```

Arguments

`elist_paths` Paths to raw illumina data files

Value

Number of numeric columns in `elist_paths` excluding probe ID columns.

iqr_replicates	<i>Removes features with replicated annotation.</i>
----------------	---

Description

For rows with duplicated annot, highested IQR retained.

Usage

```
iqr_replicates(eset, annot = "SYMBOL", rm.dup = FALSE)
```

Arguments

eset	Annotated eset created by load_raw.
annot	feature to use to remove replicates.
rm.dup	remove duplicates (same measure, multiple ids)? Used for Pathway analysis so that doesn't treat probes that map to multiple genes as distinct measures.

Value

Expression set with unique features at probe or gene level.

is_invertible	<i>Check uploaded bulk pdata to make sure the study design is invertible</i>
---------------	--

Description

Check uploaded bulk pdata to make sure the study design is invertible

Usage

```
is_invertible(pdata)
```

load_agil_plat	<i>Load Agilent raw data</i>
----------------	------------------------------

Description

Load Agilent raw data

Usage

```
load_agil_plat(eset, gse_name, gse_dir, ensql)
```

Arguments

eset	ExpressionSet from getGEO .
gse_name	Accession name for eset.
gse_dir	Direction with Agilent raw data.
ensql	For development. Path to sqlite file with ENTREZID and SYMBOL columns created in data-raw/entrezdt.

Value

ExpressionSet

load_diff	<i>Load previous differential expression analyses.</i>
-----------	--

Description

Loads previous differential expression analyses.

Usage

```
load_diff(gse_names, data_dir = getwd(), annot = "SYMBOL", postfix = NULL)
```

Arguments

gse_names	Character vector specifying GSE names to be loaded.
data_dir	String specifying directory of GSE folders.
annot	Level of previous analysis (e.g. "SYMBOL" or "PROBE").
postfix	Optional string to append to saved results. Useful if need to run multiple meta-analyses on the same series but with different contrasts.

Value

Result of previous call to [diff_expr](#).

Examples

```
library(lydata)

data_dir <- system.file("extdata", package = "lydata")
gse_names <- c("GSE9601", "GSE34817")
prev <- load_diff(gse_names, data_dir)
```

load_illum_plat	<i>Illumina loader utility for load_plat.</i>
-----------------	---

Description

Used by load_plat to load an eset.

Usage

```
load_illum_plat(eset, gse_name, gse_dir, ensql)
```

Arguments

eset	Expression set obtained by getGEO.
gse_name	String specifying GSE name.
gse_dir	String specifying path to GSE folder.

Value

Annotated eset.

See Also

[load_plat](#).

load_plat	<i>Load and pre-process raw Affymetrix, Illumina, and Agilent microarrays.</i>
-----------	--

Description

Load raw files previously downloaded with get_raw. Used by load_raw.

Usage

```
load_plat(gse_name, data_dir, gpl_dir, ensql)
```


Arguments

gse_name	GSE names.
data_dir	String specifying directory with GSE folder.
gpl_dir	String specifying parent directory to search for previously downloaded GPL.soft files.
ensql	For development. Path to sqlite file with ENTREZID and SYMBOL columns created in data-raw/entrezdt.

Details

Data is normalized, SYMBOL and PROBE annotation are added to fData slot.

Value

List of annotated esets, one for each platform in gse_name.

See Also

[get_raw](#) to obtain raw data.

load_raw	<i>Load and annotate raw data downloaded from GEO.</i>
----------	--

Description

Loads and annotates raw data previously downloaded with [get_raw](#). Supported platforms include Affymetrix, Agilent, and Illumina.

Usage

```
load_raw(
  gse_names,
  data_dir = getwd(),
  gpl_dir = "..",
  overwrite = FALSE,
  ensql = NULL
)
```

Arguments

gse_names	Character vector of GSE names.
data_dir	String specifying directory with GSE folders.
gpl_dir	String specifying parent directory to search for previously downloaded GPL.soft files.
overwrite	Do you want to overwrite saved esets from previous load_raw?
ensql	For development. Path to sqlite file with ENTREZID and SYMBOL columns created in data-raw/entrezdt.

Value

List of annotated esets.

Examples

```
library(lydata)
data_dir <- system.file("extdata", package = "lydata")
eset <- load_raw("GSE9601", data_dir = data_dir)
```

makeExampleCountsEset *Make example ExpressionSet*

Description

adapted from DESeq2::makeExampleDESeqDataSet

Usage

```
makeExampleCountsEset(
  n = 1000,
  m = 12,
  betaSD = 0,
  interceptMean = 4,
  interceptSD = 2,
  dispMeanRel = function(x) 4/x + 0.1,
  sizeFactors = rep(1, m)
)
```

Arguments

n	number of rows
m	number of columns
betaSD	the standard deviation for non-intercept betas, i.e. $\beta \sim N(0, \text{betaSD})$
interceptMean	the mean of the intercept betas (log ₂ scale)
interceptSD	the standard deviation of the intercept betas (log ₂ scale)
dispMeanRel	a function specifying the relationship of the dispersions on $2^{\text{trueIntercept}}$
sizeFactors	multiplicative factors for each sample

Examples

```
eset <- makeExampleCountsEset()
```

match_prev_eset	<i>Reuse contrast selections from previous analysis.</i>
-----------------	--

Description

Transfers user-supplied selections from previous call of `diff_expr`.

Usage

```
match_prev_eset(eset, prev_anal)
```

Arguments

<code>eset</code>	Annotated eset. Created by <code>load_raw</code> .
<code>prev_anal</code>	One item (for <code>eset</code>) from previous result of <code>diff_expr</code> . If present, previous selections and names will be reused.

Value

Expression set with samples and `pData` as in `prev_anal`.

See Also

[diff_expr](#)

<code>open_raw_illum</code>	<i>Open raw Illumina microarray files.</i>
-----------------------------	--

Description

Helper function to open raw Illumina microarray files in order to check that they are formatted correctly. For details on correct format, please see 'Checking Raw Illumina Data' in vignette.

Usage

```
open_raw_illum(gse_names, data_dir = getwd())
```

Arguments

<code>gse_names</code>	Character vector of Illumina GSE names to open.
<code>data_dir</code>	String specifying directory with GSE folders.

Value

Character vector of successfully formatted Illumina GSE names.

Examples

```
library(lydata)

# Illumina GSE names
illum_names <- c("GSE50841", "GSE34817", "GSE29689")

# location of raw data
data_dir <- system.file("extdata", package = "lydata")

# open raw data files with default text editor
# open_raw_illum(illum_names)
```

phenoData.ch2

Construct AnnotatedDataFrame from Two-Channel ExpressionSet

Description

Construct AnnotatedDataFrame from Two-Channel ExpressionSet

Usage

```
phenoData.ch2(eset)
```

Arguments

eset ExpressionSet with pData for two-channel Agilent array.

Value

AnnotatedDataFrame with twice as many rows as eset, one for each channel of each array in order all red then all green.

prefix_illum_headers *Run prefix on Illumina raw data files*

Description

Run prefix on Illumina raw data files

Usage

```
prefix_illum_headers(elist_paths)
```

Arguments

elist_paths Paths to raw Illumina data files

Value

Paths to fixed versions of `elist_paths`

query_ref	<i>Get correlation between query and reference signatures.</i>
-----------	--

Description

Determines the pearson correlation between the query and each reference signature.

Usage

```
query_ref(query, ref, sorted = TRUE, ngenes = 200)
```

Arguments

query	Named numeric vector of differential expression values for query genes. Usually 'meta' slot of <code>get_dprimes</code> result.
ref	A matrix of differential expression to query against (rows are genes, columns are samples).
sorted	Would you like the results sorted by decreasing similarity? Default is TRUE.
ngenes	The number of top differentially-regulated (up and down) query genes to use.

Value

Vector of pearson correlations between query and reference signatures.

remove_autonamed	<i>Remove columns that are autonamed by data.table</i>
------------------	--

Description

Auto-named columns start with 'V' followed by the column number.

Usage

```
remove_autonamed(ex)
```

Arguments

ex	data.frame loaded with fread
----	--

Value

ex with auto-named columns removed.

run_limma	<i>Linear model fitting of eset with limma.</i>
-----------	---

Description

After selecting control and test samples for a contrast, surrogate variable analysis ([sva](#)) and linear model fitting with [lmFit](#) is performed.

Usage

```
run_limma(  
  eset,  
  annot = "SYMBOL",  
  svobj = list(sv = NULL),  
  numsv = 0,  
  filter = TRUE  
)
```

Arguments

eset	Annotated eset created by load_raw .
annot	String, column name in fData. For duplicated values in this column, the row with the highest interquartile range across selected samples will be kept. Appropriate values are "SYMBOL" (default - for gene level analysis) or "ENTREZID_HS" (for probe level analysis).
svobj	Surrogate variable analysis results. Returned from run_sva .
numsv	Number of surrogate variables to model.
filter	For RNA-seq. Should genes with low counts be filtered? dseqr shiny app performs this step separately. Should be TRUE (default) if used outside of dseqr shiny app .

Details

If analyses need to be repeated, previous results can be reloaded with [readRDS](#) and supplied to the `prev_anal` parameter. In this case, previous selections will be reused.

Value

List with:

fit	result of lmFit .
mod	<code>model.matrix</code> used for fit

run_limma_setup	<i>Setup ExpressionSet for running limma analysis</i>
-----------------	---

Description

Setup ExpressionSet for running limma analysis

Usage

```
run_limma_setup(eset, prev)
```

Arguments

eset	ExpressionSet
prev	previous result of call to diff_expr

Value

eset ready for run_limma

run_lmfit	<i>Perform lmFit analysis from limma.</i>
-----------	---

Description

If paired samples, runs [duplicateCorrelation](#) to estimate intra-patient variance.

Usage

```
run_lmfit(eset, mod, rna_seq = TRUE)
```

Arguments

eset	Annotated eset created by load_raw. Non-selected samples and duplicate features removed by add_contrasts and iqr_replicates.
mod	Model matrix generated by diff_setup. With or without surrogate variables.
rna_seq	Is this an RNA-seq eset? Default is TRUE.

Value

result from call to limma lmFit.

run_select_contrasts *Shiny gadget to upload groups and select contrasts*

Description

Shiny gadget to upload groups and select contrasts

Usage

```
run_select_contrasts(
  eset,
  gse_name,
  prev = NULL,
  app_dir = system.file("select_contrasts", package = "crossmeta", mustWork = TRUE),
  port = 3838
)
```

Arguments

eset	ExpressionSet
gse_name	GEO accession for the series.
prev	Previous result of diff_expr. Used to allow rechecking previous selections.
app_dir	Directory to shiny app. For local development use 'inst/select_contrasts'. Default is in 'select_contrasts' sub directory of crossmeta package.
port	See runApp() .

Value

result of [setup_prev](#). Used to specify sample groups and contrasts for differential expression analysis.

run_sva *Run surrogate variable analysis*

Description

Run surrogate variable analysis

Usage

```
run_sva(mods, eset, svanal = TRUE)
```


Arguments

mods	result of get_sva_mods
eset	ExpressionSet
svanal	Should surrogate variable analysis be run? If FALSE, returns dummy result.

setup_prev	<i>Setup selections when many samples.</i>
------------	--

Description

Function is useful when number of samples makes manual selection with [diff_expr](#) error prone and time-consuming. This is often true for large clinical data sets.

Usage

```
setup_prev(eset, contrasts)
```

Arguments

eset	List containing one expression set with pData 'group' and 'pair' (optional) columns. Name of eset should be the GSE name.
contrasts	Character vector specifying contrasts to analyse. Each contrast must take the form "B-A" where both "B" and "A" are present in eset pData 'group' column. "B" is the treatment group and "A" is the control group.

Value

List containing necessary information for prev_anal parameter of [diff_expr](#).

Examples

```
library(lydata)
library(Biobase)

# location of raw data
data_dir <- system.file("extdata", package = "lydata")

# load eset
gse_name <- c("GSE34817")
eset <- load_raw(gse_name, data_dir)

# inspect pData of eset
# View(pData(eset$GSE34817)) # if using RStudio
head(pData(eset$GSE34817)) # otherwise

# get group info from pData (differs based on eset)
group <- pData(eset$GSE34817)$characteristics_ch1.1
```

```
# make group names concise and valid
group <- gsub("treatment: ", "", group)
group <- make.names(group)

# add group to eset pData
pData(eset$GSE34817)$group <- group

# setup selections
sel <- setup_prev(eset, contrasts = "LY-DMSO")

# run differential expression analysis
anal <- diff_expr(eset, data_dir, prev_anal = sel)
```

symbol_annot	<i>Add hgnc symbol to expression set.</i>
--------------	---

Description

Function first maps entrez gene ids to homologous human entrez gene ids and then to hgnc symbols.

Usage

```
symbol_annot(eset, gse_name = "", ensql = NULL)
```

Arguments

eset	Expression set to annotate.
gse_name	GSE name for eset.
ensql	For development. Path to sqlite file with ENTREZID and SYMBOL columns created in data-raw/entrezdt.

Details

Initial entrez gene ids are obtained from bioconductor annotation data packages or from feature data of supplied expression set. Homologous human entrez ids are obtained from homogene and then mapped to hgnc symbols using org.Hs.eg.db. Expression set is expanded if 1:many mappings occur.

Value

Expression set with hgnc symbols ("SYMBOL") and row names ("PROBE") added to fData slot.

See Also

[load_raw](#).

Examples

```
library(lydata)

# location of raw data
data_dir <- system.file("extdata", package = "lydata")

# load eset
eset <- load_raw("GSE9601", data_dir)[[1]]

# annotate eset (need if load_raw failed to annotate)
eset <- symbol_annot(eset)
```

to_eset	<i>Convert limma object to ExpressionSet</i>
---------	--

Description

Convert limma object to ExpressionSet

Usage

```
to_eset(object, eset)
```

Arguments

object	an EList of MAList object containing expression data.
eset	ExpressionSet from getGEO . Used for annotation.

Value

ExpressionSet using expression data from object and annotation from eset.

to_ma	<i>Covert expression values to MAList</i>
-------	---

Description

Covert expression values to MAList

Usage

```
to_ma(y)
```

Arguments

y	Expression values from two-channel agilent array in order all red then all green.
---	---

Value

MAList

Examples

```
A <- matrix(rnorm(100), ncol = 5)
M <- matrix(rnorm(100), ncol = 5)
MA <- new('MAList', list(M=M, A=A))
colnames(MA) <- letters[1:5]
```

```
y <- exprs.MA(MA)
MA2 <- crossmeta:::to_ma(y)
all.equal(MA, MA2)
```

validate_up_annot	<i>Validate uploaded bulk annotation</i>
-------------------	--

Description

Validate uploaded bulk annotation

Usage

```
validate_up_annot(up, ref)
```

which_max_iqr	<i>Get row indices of maximum IQR within annotation groups</i>
---------------	--

Description

Groups by group_by and determines row with maximum IQR.

Usage

```
which_max_iqr(eset, group_by, x = exprs(eset))
```

Arguments

eset	ExpressionSet
group_by	Column in fData(eset) to group by
x	matrix of expression values to use for IQR

Value

Integer vector of row numbers representing rows with the maximum IQR after grouping by group_by

xls_to_txt	<i>Covert .xls files to .txt</i>
------------	----------------------------------

Description

For converting Illumina _Supplementary_*.xls files to .txt for load_illum_plat.

Usage

```
xls_to_txt(xls_paths)
```

Arguments

xls_paths	Paths to .xls files
-----------	---------------------

Index

- * <-
 - run_select_contrasts, 32
- * =
 - run_select_contrasts, 32
- * **GSE**
 - run_select_contrasts, 32
- * #
 - run_select_contrasts, 32
- * **analysis**
 - run_select_contrasts, 32
- * **data_dir**[[1]]
 - run_select_contrasts, 32
- * **data_dir**
 - run_select_contrasts, 32
- * **datasets**
 - gs.names, 20
 - gslist, 21
- * **data**
 - run_select_contrasts, 32
- * **eset**
 - run_select_contrasts, 32
- * **gather**
 - run_select_contrasts, 32
- * **gse_name**
 - run_select_contrasts, 32
- * **gse_name**
 - run_select_contrasts, 32
- * **internal**
 - addContrastInput, 3
 - bulkAnnot, 6
 - bulkAnnotInput, 6
 - bulkForm, 6
 - bulkFormInput, 7
 - bulkTable, 8
 - bulkTableOuput, 8
 - ch2_subset, 8
 - clean_y, 9
 - delContrastsInput, 9
 - fit_lm, 15
 - format_dl_annot, 16
 - format_up_annot, 16
 - get_ch2_mod, 16
 - get_group_levels, 17
 - get_palette, 17
 - is_invertible, 22
 - load_illum_plat, 24
 - load_plat, 24
 - match_prev_eset, 27
 - query_ref, 29
 - run_lmfit, 31
 - run_select_contrasts, 32
 - to_eset, 35
 - to_ma, 35
 - validate_up_annot, 36
 - which_max_iqr, 36
 - xls_to_txt, 37
- * **library(lydata)**
 - run_select_contrasts, 32
- * **load_raw(gse_name,**
 - run_select_contrasts, 32
- * **load**
 - run_select_contrasts, 32
- * **location**
 - run_select_contrasts, 32
- * **lydata)**
 - run_select_contrasts, 32
- * **names**
 - run_select_contrasts, 32
- * **of**
 - run_select_contrasts, 32
- * **package**
 - run_select_contrasts, 32
- * **previous**
 - run_select_contrasts, 32
- * **run_select_contrasts(eset,**
 - run_select_contrasts, 32
- * **system.file(extdata,**
 - run_select_contrasts, 32

* **z**
 run_select_contrasts, 32

add_adjusted, 4
add_sources, 4, 12
add_vsd, 5
addContrastInput, 3

bulkAnnot, 6
bulkAnnotInput, 6
bulkForm, 6
bulkFormInput, 7
bulkPage, 7
bulkPageUI, 7
bulkTable, 8
bulkTableOutput, 8

ch2_subset, 8
clean_y, 9
contrasts.fit, 14, 19

delContrastsInput, 9
diff_expr, 4, 5, 10, 10, 12, 23, 27, 33
duplicateCorrelation, 31

eBayes, 11, 14, 19
effectsize, 12, 19
es_meta, 11
exprs.MA, 13, 13

fdrtool, 12
filter_genes, 13
filterByExpr, 13, 14
fit_ebayes, 14
fit_lm, 15
fix_illum_headers, 15
format_dl_annot, 16
format_up_annot, 16
fread, 29

get_ch2_mod, 16
get_group_levels, 17
get_palette, 17
get_raw, 5, 11, 18, 25
get_sva_mods, 18, 33
get_top_table, 19
get_vsd, 20
getGEO, 15, 23, 35
gs.names, 20
gslist, 21

ilmn.nnum, 21
intraspotCorrelation, 16
iqr_replicates, 22
is_invertible, 22

lmFit, 15, 30
lmscFit, 16
load_agil_plat, 23
load_diff, 4, 10–12, 23
load_illum_plat, 24
load_plat, 24, 24
load_raw, 10, 18, 20, 25, 34

makeExampleCountsEset, 26
match_prev_eset, 27

open_raw_illum, 27

phenoData.ch2, 13, 28
plotDensities.MAList, 13
prefix_illum_headers, 28

query_ref, 29

read.ilmn, 15
readRDS, 30
remove_autonamed, 29
run_limma, 14, 19, 30
run_limma_setup, 31
run_lmfit, 31
run_select_contrasts, 32
run_sva, 15, 30, 32
runApp(), 10, 32

setup_prev, 32, 33
sva, 10, 30
symbol_annot, 34

to_eset, 35
to_ma, 35
topTable, 11
toptable, 19

validate_up_annot, 36

which_max_iqr, 36

xls_to_txt, 37

zScores, 12